

Effects of Consumption of Various Formulations of Cocoyam-Bambara Groundnut-Soya Bean Flour Blends on the Pancreatic Beta-Cell Functions of Streptozotocin-Induced Diabetic Rats

Uro-Chukwu HC^{1,2,3*}, Ezekwe AS⁴, Okari KA⁴, Oko IC⁵, Uro-Chukwu FNC² and Uro-Chukwu FCU²

¹Department of Medical Biochemistry, Alex Ekwueme Federal University, Ndufu-Alike, Ikwo, Nigeria

²Institute for Nutrition, Nutraceuticals & Public Health Research & Development, Nigeria

³Department of Community Medicine, Ebonyi State University, Abakaliki, Nigeria

⁴Department of Medical Biochemistry, Rivers State University, Nkporlu, Port Harcourt, Nigeria

⁵University of Glasgow, United Kingdom

*Corresponding author

Uro-Chukwu, Henry Chukwuemeka, Department of Medical Biochemistry, Alex Ekwueme, Federal University, Ndufu-Alike, Ikwo, Nigeria.

Received: January 15, 2025; **Accepted:** January 27, 2025; **Published:** February 03, 2025

Keywords: HOMA-IR, HOMA- β , Medicinal Plant Foods, Diabetic Rats

Introduction

Diabetes mellitus is an endocrine system disease affecting people of diverse race, ages and regions globally, increasing in number annually due to its long course and recalcitrant to conventional treatment [1]. Its clinical presentations are multi-system based, affecting virtually all key organs and which as a result of the chronicity of its course ends in organ failures [2]. The disease affects more people in economically disadvantaged countries compared to economically buoyant countries [3]. This high prevalence in low-income nations has been attributed to the weak health systems, dearth of health personnel, modern equipment, infrastructure and disappointing management outcomes [4-6]. In finding solutions to such negative outcomes, management protocols have been expanded to target lifestyle modifications including focus on dietary plan and increased physical activity levels [7]. Consequently food-based plans have been incorporated as immediate and long term critical component in T2DM management, the positive results of which have made global experts and bodies, including the World Health Organization to encourage adoption of medicinal plant foods in T2DM treatment and prophylaxis [8-10]. The major scientific basis for utilization of medicinal plant foods is its composite bioactive compounds with reported biological *in vivo* activities [11,12]. These bio-actions ranges from potent effects against free

radicals, its actions against inflammation, the pro-hypoglycemic effects and their activities that boost the immune system [11,12]. Bioactive compounds associated with these bio-actions include low-molecular weight compounds like peptides, amino acids, and other biological agents such as flavonoids, and polyphenols, all of which acting on the pancreatic β -cells and involved in the hepatic glycol-regulatory enzymes [13]. In the developing countries, commonly consumed plant foods such as the roots of Cocoyam, have been demonstrated to contain biologically active substances with established anti-hyperglycemic effects [14-16]. Legumes like Bambara groundnut (*Vigna subterranean*), also contain several biologically active compounds in addition to its nutrients [17-26]. These compounds found in bambara groundnut use various mechanisms of actions to effect hypoglycemic actions [27-31]. Another ubiquitous legume, Soya Bean (*Glycine max.* (L) Merrill), also has abundant biologically active compounds responsible for beneficial effects when incorporated in the management of diabetic mellitus [32-36]. Most of these mechanisms of actions have been adequately reported from studies [37-46]. Since these plant foods are usually consumed in combinations, this study intended to evaluate the effect of these flour blends in the pancreatic tissues of diabetic rats.

Collection and Preparation of the Plant Food Material

Locally sourced soy bean seeds, roots of cocoyam and seeds of Bambara groundnut were processed into pellets by first washing them, peeling the cocoyam, and soaking each in water for 10

min, then rinsing and boiling them. They were later oven-dried at a temperature of 60°C. Once the weights of the products became constant, they were ground into fine flour, pelletized, oven-dried again and stored for subsequent use in a well-sealed container.

Experimental Animals

A total of Eighty-five (85) male albino rats whose total body weights ranged from 134 - 249 grams were purchased from a reputable breeder. The rats were separated into groups of eight with each rat marked on the tail, using an indelible, and the marks depicting its number (1 - 8). In all, ten sets of the rats, A-J, were used in the experiment, with the weight of each rat in the set, documented, every week. All the rats were fed with commercial rat feeds and water for one week ad libitum to allow for acclimatization. The standard recommended protocol for administration of laboratory animals in experiment were strictly followed [47].

Induction of Insulin Resistance Using Low Fructose Diet

After acclimatizing the experimental animals for one week, a low-dose fructose diet was instituted to replace the drinking water, by administering the rats with 300 mls of 10% fructose ad libitum, in order to induce Insulin resistance. The commercial rat feeds were equally administered ad libitum within this period.

Induction of Type 2 Diabetes Mellitus Using STZ

The 10% fructose diet is administered for 6 days at the end of which the rats are fasted overnight and the fasting blood glucose level measured. Rats that were normoglycemic were then administered with Streptozotocin (STZ) which was prepared fresh by dissolving 1g of STZ in 50 mL of freshly prepared sodium citrate buffer, 0.1M, pH 4.5), at the recommended dose for extracts in experimental animals [48,49]. Following the administration of the STZ, the rat blood samples were collected for blood glucose level evaluation, at the tail veins at specified periods of 72 hours and on day 12. All the rats whose blood glucose levels were hyperglycemic were then randomly assigned to different groups: the standard control, negative control, and seven intervention groups. The rats who were not treated to the STZ and which remained normoglycemic, were assigned as the normal control group. Common to all the ten rat groups was the administration of commercial rat feeds for 28 days, but the specific rat groups were additionally given metformin at 200 mg/kg, as 0.002 ml per Kg body weight orally daily using oral dispenser (standard control), formulations 1, 2, 4, 7, 5, 6 and 3 for rat groups A, E, F, G, H, I and J respectively, according to the work of Nnadi and colleagues (Table 1) [50].

Table 1: The Formulations and Their Compositions Given to Each Rat Group

Rat Groups	Formulations	Formulation Compositions
A	1	16.6%CY: 16.6%SB: 16.6%BGN: 50%RF
B	Standard Control	Metformin + 100% Rat Feed (RF)
C	Normal Control	100% RF
D	Negative Control	100% RF
E	2	12.5%CY: 12.5%SB: 25%BGN: 50%RF
F	4	12.5%CY: 25%SB: 12.5%BGN: 50%RF
G	7	0%CY: 50%SB: 0%BGN: 50%RF
H	5	0%CY: 0%SB: 50%BGN: 50%RF
I	6	50%CY: 0%SB: 0%BGN: 50%RF
J	3	25%CY: 12.5%SB: 12.5%BGN: 50%RF

RF = Rat Feed

CY = Cocoyam

SB = Soya Bean

BGN = Bambara Groundnut

Estimation of Blood Glucose and Insulin Concentrations

The interventions in each of the rats in groups A - J, were conducted for a total of 28 days, with the random and fasting blood glucose levels checked weekly. At the end of 28 days, the rats were fasted overnight and the fasting blood glucose and insulin concentrations, measured.

Statistical Analysis

The Statistical Package for Social Sciences (SPSS Inc., Chicago, IL) version 20.0 was used to analyze the data so obtained. The data were expressed as the standard error of the mean (SEM) of the triplicate values for each of the rats per group and the mean recorded. The mean difference was determined using Turkey's post-hoc test with the level of statistical significance put at P<0.05. The means were compared using a one-way analysis of variance (ANOVA).

Results

Random Blood Glucose Concentration, mg/dl, (Weeks 1-4) of the Various Groups of STZ-Induced Diabetic and Non-Diabetic Rats.

Table 2: Showing Result of the Random Blood Glucose Concentration, mg/dl, (Weeks 1-4) of the Various Groups of STZ-Induced Diabetic and Non-Diabetic Rats

Groups	Week 1 Mean (SEM)	Week 2 Mean (SEM)	Week 3 Mean (SEM)	Week 4 Mean (SEM)
A (n=7)	315.83(22.23) ^b	396.33(11.61) ^{bc}	457.67(24.91) ^b	512.67(8.67) ^b
B(n=7)	401.50(13.98) ^{bc}	309.00(48.32) ^b	371.00(67.06) ^b	414.25(75.15) ^b
C(n=9)	106.25(3.05) ^a	101.00(6.22) ^a	112.88(4.25) ^a	94.13(1.49) ^a
D(n=5)	395.67(30.98) ^{bc}	332.33(34.61) ^{bc}	364.00(15.36) ^b	563.00(11.32) ^b
E(n=7)	400.20(23.02) ^{bc}	320.60(21.43) ^b	392.20(25.30) ^b	526.80(14.65) ^b
F(n=7)	365.33(17.12) ^{bc}	356.67(9.49) ^{bc}	410.67(10.46) ^b	542.67(20.15) ^b

G(n=7)	367.86(49.08) ^{bc}	326.43(42.40) ^{bc}	379.43(45.85) ^b	457.43(59.59) ^b
H(n=7)	362.00(17.66) ^{bc}	291.20(48.44) ^b	313.60(59.36) ^b	481.60(45.55) ^b
I(n=7)	434.00(14.55) ^c	448.33(32.06) ^c	453.00(23.95) ^b	497.67(3.78) ^b
J(n=7)	367.17(13.60) ^{bc}	366.50(11.91) ^{bc}	403.00(6.82) ^b	481.83(12.17) ^b
F	14.10	7.10	6.41	10.89
p-value	<0.001	<0.001	<0.001	<0.001

Repeated Measures Test

Equal Level (Mean difference among Groups)		Parallelism (Pattern of all groups across time points) i.e. within/between group interactions		Flatness (Trend)	
F-value	p-value	F-value	p-value	F-value	p-value
13.963	<0.001	79.106	<0.001	76.755	<0.001

The Table 2, The ANOVA result depicting blood glucose concentration mean differences among groups A-J and as well as determining the trend over time are captured in Table 2, revealing the significant difference in the mean over time among the groups (F= 13.963, p<0.001), the parallel pattern of changes over time which was also significant (F=79.106, p<0.01) as well as the significant positive trend (F=76.755, p<0.001).

Table 3: Showing the Result of the Fasting Blood Glucose (mg/dl) (Day of Sacrifice) of the various Groups of STZ-Induced Diabetic and Non-Diabetic Rats

Group	Mean	Standard Error of Mean	Minimum	Maximum
A (n=7)	294.57 ^b	44.94	75	416
B(n=7)	282.25 ^{ab}	64.62	89	360
C(n=9)	76.38 ^a	2.63	65	85
D(n=5)	338.67 ^b	10.51	324	359
E(n=7)	286.20 ^{ab}	36.71	170	395
F(n=7)	361.14 ^b	20.96	291	422
G(n=7)	318.00 ^b	43.35	81	416
H(n=7)	312.40 ^b	56.54	89	405
I(n=7)	296.67 ^b	138.12	96	561
J(n=7)	241.00 ^{ab}	38.37	110	318
F	4.38			
p-value	<0.001			

Table 3 captured the mean fasting blood glucose concentrations for the various Groups with Groups C and F showing the least and highest values respectively, Groups E, and J were not significantly different from the standard control.

Figure 1 is the Pareto bar for the insulin concentrations depicting the highest mean value from the standard control and the least value from group A. With the exception of Group B, all the group means were not statistically different from one another.

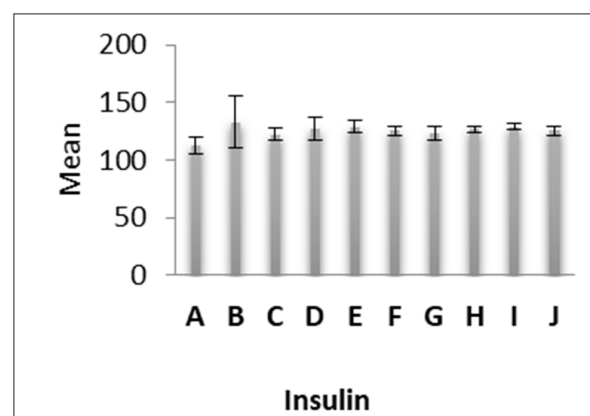


Figure 1: Pareto Bar Chart of Insulin

Table 4: Calculated HOMA-IR and HOMA-β Values for diabetic and non-diabetic Rat Groups

Groups	Fasting Blood Glucose (mg/dl) (Day of Sacrifice)	Fasting Plasma Insulin (mIU/mL) Mean(SEM)	HOMA-IR	HOMA-β
A	294.571 ^b	112.49(2.79) ^a	0.82	1.75
B	282.250 ^{ab}	133.17(8.31) ^b	0.94	2.18
C	76.375 ^a	122.45(1.95) ^{ab}	0.23	32.83
D	338.667 ^b	127.43(3.74) ^{ab}	1.06	1.66
E	286.200 ^{ab}	128.83(2.01) ^b	0.9	2.08
F	361.143 ^b	125.33(1.49) ^{ab}	1.11	1.51
G	318.000 ^b	122.84(2.29) ^{ab}	0.97	1.74
H	312.400 ^b	126.68(1.49) ^{ab}	0.98	1.83
I	296.667 ^b	128.80(1.00) ^b	0.94	1.99
J	241.000 ^{ab}	125.66(1.51) ^{ab}	0.75	2.55

Table 4 shows the calculated values of HOMA-IR and HOMA- β from the study results of the fasting blood glucose and fasting blood insulin on the day of sacrifice of the non-diabetic and STZ-induced diabetic rats. While the HOMA-IR reports in all the groups were within the normal range with those of groups D and F recording the highest values and tending towards the upper limits of normal, the HOMA- β values were higher than the controls signifying a decrease in the pancreatic β -cell function possibly due to the hyperglycemic effects and toxicity of the STZ.

Discussion

Effects on Blood Glucose Concentrations

Over the four weeks period, the random blood glucose (RBG) values showed significant differences for each group ($F=13.963$, $p<0.001$). These differences had similar pattern for the whole group and such differences were significantly parallel ($F=79.106$, $p<0.01$), even in terms of the time trend ($F=76.755$, $p<0.001$). There were two distinctive groups, Group C, Group B and Group D, representing the control, standard and negative groups respectively in ascending order of values on one hand, and the intervention groups E,F,G and H which had low RBG values and which when compared with the standard and diabetic control groups showed lower RBG values, implying that the formulations 2, 4, 7 and 5 respectively used in these groups (Table 1) regulated the blood glucose better than the RF used in the diabetic control and the metformin used in the standard control.

The Fasting Blood Glucose (FBG) measured after an overnight fast showed that the standard control, Group E and J had values which were not significantly different from one another but higher and significantly different from the normal control (Table 3). FBG is a better indicator of diabetic control than RBG.

It is possible that constituents like the bioactive compounds seen in the formulations 2 and 3 used for groups E and J respectively, had better efficacy when compared with metformin used in the standard control. The variations in FBG values observed over the time trend are similar to therapeutic responses observed human diabetics, hence the use of multiple anti-diabetic drug regimen [51,52]. Among the BACs found in formulations 2 and 3 are phenolic compounds which causes hypoglycemia by acting on 5' adenosine monophosphate-activated protein kinase (AMPK) pathway, improving glucose metabolism enhancing the α -amylase and α -glucosidase inhibitory activities [53-55]. Many studies have associated presence of phenolic compounds with hypoglycaemia [56-58].

Monoterpenoids have also been reported to be present in formulation 7 given to group G (53) and their derivatives exert inhibitory actions on some enzymes to effect hypoglycaemic effects [59,60].

Rats in Group I who received formulation 6, might have had artemisinin compounds whose derivatives have been associated with hypoglycaemic effects as a result of their inhibitory actions on α -glucosidase activity [61]. Other invivo action of the derivatives of artemisinin are improvement of insulin release and its actions on the pancreas [62-64]. Studies have linked artemisinin with amelioration of diabetic complications [65]. Various other studies detailing several mechanisms of action have given boost to the antidiabetic effects of artemisinin [63,65-69].

Another bioactive compound found in formulation 2 and the commercial rat feed (RF) is the Thiadiazole [53]. The chemical structure of thiadiazole lend credence to their anti-diabetic effects and that of their derivatives through their inhibitory actions on carbonic anhydrase [70-73]. Formulation 2 was the intervention formulation given to rats in group E and the FBG values measured in those groups of diabetic rats, depicted similarity with values obtained from the standard control in this work (Tables 2 & 3).

Thiourea and Naphthalene were found in formulations 4 and 7, respectively, with the former already in use for its various therapeutic effects especially too in the synthesis of sulfonylureas for diabetic management and with their derivatives acting through different mechanisms to achieve hypoglycemic effects [74-76]. Similarly, naphthalene inhibits Fatty acid binding protein 4 (FABP4), besides exerting peroxisome proliferator-activated receptor gamma (PPAR γ) agonistic activity, all of which have been implicated in fasting blood glucose reduction [64,77]. The rats in Group G were fed with formulation 7, and exhibited lower FBG than in Group D rats.

Effect on Pancreatic Cell Function and Heart Disease Risk

Insulin resistance (IR) and pancreatic β -cell function have been linked with the development of T2DM. Homeostatic Model Assessment (HOMA) has been used to assess both the IR and pancreatic β -cell function because it reveals the quantity of functional insulin, the pancreas should produce to control the blood sugar level [78,79]. Decreased pancreatic β -cell function in an existing IR is an important determinant of T2DM incidence and advancement [80]. In this study, the HOMA IR and HOMA- β were calculated using the formula of Katsuki and colleagues, with glucose in mass unit mg/dl as follows [81]:

$$\text{HOMA-IR} = \text{Glucose} \times \text{Insulin} \% / 405$$

$$\text{HOMA-} \beta = 360 \times \text{Insulin} \% / \text{Glucose} - 63$$

The values obtained (Table 4) were interpreted using the standard blood code (thebloodcode.com), while for HOMA- β , higher levels is assumed to be protective of the pancreatic cell function but lower values signify non-protection. The more significant the IR, the higher indication of the heart disease risk.

All the groups in this study, when compared to normal control, had higher HOMA-IR values (Table 4). This documented report was in tandem with some other works which reported that subjects with IR and T2DM, had higher HOMA-IR than controls [82,83]. The HOMA-IR values in this study were between 0.23 - 1.11, which were all within the normal limit of 0.5 - 1.9, meaning that the formulations and the antidiabetic drug used in the study, exerted a balance in the dynamics between the FBG and the responsive insulin, thereby limiting the development of IR. Groups D and F had the highest values of 1.06 and 1.11 respectively, which tended towards the upper limit of normal, depicting a reduced potency of formulation 4 (12.5%CY: 25%SB: 12.5%BGN: 50%RF) used for group F and the non-protective nature of non-intervention in the diabetic control (Table 4).

On the HOMA- β , the study showed that all the groups had high values with group C exhibiting the highest value, while group F and the diabetic control, had the least. High HOMA- β

is an indication of protection of pancreatic- β cells and hence against T2DM [84]. The documented reports from this study were not in tandem with the report by Sung and colleagues, since the HOMA- β in all the groups were far lower than the value of 32.83 obtained in the normal control (Table 4). Previous studies have reported that low HOMA- β levels were predictive of T2DM [85,86]. HOMA calculations is based on a computer-solved model that assumes certain relationship between basal plasma glucose and insulin concentration [86,87]. Using reports from the values of the HOMA- β in this study, it can be inferred that there was a reduction in the pancreatic β -cells with the administration of the STZ and the resultant hyperglycemia but the administration of the intervention formulations might have aided the regeneration of the beta cells which histological studies showed and which other reports had documented [88-90].

Conclusions

Flour blend formulations from cocoyam-soya bean-Bambara groundnut, when administered to SDRs in various ratios exerted improvement in the pancreatic β -cell functions and limited the advancement of insulin resistance and development of T2DM in SDRs, and these actions were comparable to the effect of metformin, anti-diabetic drug. Among the various blend formulations, formulation 4 (12.5%CY: 25%SB: 12.5%BGN: 50%RF) had the least potency, while formulation 3 (25%CY: 12.5%SB: 12.5%BGN: 50%RF) had the highest potency in HOMA- β . The implication of this study is that the consumption of cocoyam-soya bean- bamabara groundnut food mix can be considered beneficial to people at risk of developing T2DM and hence recommended as a nutritional management support.

Authors' Contributions:

HU =Topic design, Proposal writing, Literature search, draft manuscript

AE = Topic design, Statistical analysis, draft manuscript

FR = Study Analysis, Literature search

KO = Manuscript writing

FU = Data Entering and literature search

All authors read and approved the final manuscript

Competing Interest

There is no competing interest.

Acknowledgement

Authors wish to acknowledge the technical support of Dr. Eleazu Chinedu and Dr. Okorie, Uchechukwu for their technical support during the experiment.

References

1. Sun H, Saeedi P, Karuranga S, Pinkepank M, Ogurtsova K, et al. IDF Diabetes Atlas: Global, Regional and Country-level diabetes prevalence estimates for 2021 and projections for 2045. *Diabetes Research and Clinical Practice*. 2022. 183: 109119.
2. VanBelle TL, Coppieters KT, Von Herrath MG. Type 1 diabetes: etiology, immunology and therapeutic strategies. *Physiological Reviews*. 2011. 91: 1.
3. Lin X, Xu Y, Pan X, Xu J, Ding Y, et al. Global, regional, and national burden and trend of diabetes in 195 countries and territories: an analysis from 1990 to 2025. *Sci Rep*. 2020. 10: 14790.
4. Kirigia JM, Sambo HB, Sambo LG, Barry SP. Economic burden of diabetes mellitus in the WHO African region. *BMC International Health and Human Rights*. 2009. 9: 6.
5. Ashcroft FM, Ashcroft SJH. *Insulin, Molecular Biology to Pathology*. Oxford University Press. 1992. 266-284.
6. Sobngwi E, Mauvais-Jarvis F, Vexiau P, Mbanya JC, Gautier JF. Diabetes in Africans. *Epidemiology and clinical specificities*. *Diabetes Metab*. 2001. 27: 628-634.
7. Piero MN. Hypoglycemic effects of some Kenyan plants traditionally used in management of diabetes mellitus in eastern province, Msc thesis, Kenyatta University. 2006.
8. Wylie-Rosett J, Delahanty LM. *Nutrition in the Prevention and Treatment of Disease*. 2017. 691-707.
9. Forouhi GN, Misra A, Mohan V, Taylor R, Yancy W. Dietary and nutritional approaches for prevention and management of type 2 Diabetes, *Science and Politics of Nutrition*. *BMJ*. 2018. 361.
10. Piero MN, Njagi JM, Kibiti CM, Ngeranwa JJN, Njagi ENM, et al. The Role of Vitamins and Mineral Elements in Management of Type 2 Diabetes Mellitus: A Review South As. *J Biol Sci*. 2012. 2: 107-115.
11. Merida LA, Mattos EB, Correa AC, Pereira PR, Paschoalin VM, et al. Tarin stimulates granulocyte growth in bone marrow cell cultures and minimizes immunosuppression by cyclo-phosphamide in mice. *plos one*. 2018. 13: e0206240.
12. Chukwuma CI, Islam MS, Amonsou EO. A comparative study on the physicochemical, anti-oxidative, anti-hyperglycemic and anti-lipidemic properties of amadumbe (*Colocasia esculenta*) and okra (*Abelmoschus esculentus*) mucilage. *j food biochem*. 2018. 42: e12601.
13. Prabhakar PK, Doble M. A Target Based Therapeutic Approach Towards Diabetes Mellitus Using Medicinal Plants. *Current Diabetes Reviews*. Bentham Science Publishers Ltd. 2008. 291-308.
14. Kumawat NS, Chaudhari SP, Wani NS, Deshmukh TA, Patil VR. Antidiabetic activity of ethanol extract of *Colocasia esculenta* leaves in alloxan induced diabetic rats. *International Journal of Pharmaceutical Technique Research*. 2010. 2: 1246-1249.
15. Eleazu CO, Iroaganachi M, Eleazu KC. Ameliorative potentials of cocoyam (*Colocasia esculenta* L.) and unripe plantain (*Musa paradisiaca* L.) on the relative tissue weights of streptozotocin-induced diabetic rats. *J Diabetes Res*. 2013. 1-8.
16. Uro-Chukwu HC, Ezekwe AS, Roberts F, Okari KA, Uro-Chukwu FNC. Anti-hyperglycemic Effects and Histological Changes in Streptozotocin-induced diabetic rats fed with high concentration of *Colocasia esculenta* flour. *south asian research journal of natural products*. 2024. 7: 123-134.
17. Arise AK, Amonsou EO, Ijabadeniyi OA. Influence of extraction methods on functional properties of protein concentrates prepared from South African Bambara groundnut landraces. *International Journal of Food Science and Technology*. 2015. 50: 1095-1101.
18. Oyeyinka SA, Oyeyinka AT. A review on isolation, composition, physicochemical properties and modification of Bambara groundnut starch. *food hydrocolloid*. 2018. 75: 62-71.
19. Oyeyinka SA, Tijani ST, Oyeyinka AT, Arise AK, Balogun MA, et al. Value added snacks produced from Bambara groundnut (*Vigna subterranea*) paste or flour. *lwt-food science & technology*. 2018. 88: 126-131.

20. Halimi RA, Barkla BJ, Mayes S, King GJ. The potential of the underutilized pulse bambara groundnut (*Vigna subterranea* (L.) Verdc.) for nutritional food security. *The Journal of Food Composition and Analysis*. 2019. 77: 47-59.
21. Olaleye AA, Adeyeye EI, Adesina AJ. Chemical Composition of Bambara groundnut (*Vigna subterranea* L. Verdc) Bangladesh Journal of Scientific & Industrial Research. 2013. 48: 167-178.
22. Salawu. Comparative study of the antioxidant activities of methanolic extract and simulated gastrointestinal enzyme digest of Bambara nut (*Vigna subterranean*) FUTA. *J Res Sci*. 2016. 1: 107-120.
23. Harris T, Jideani V, Le Rose-Hill M. Flavonoids and tannin composition of Bambara groundnut (*Vigna subterranean*) of Mpumalanga, South Africa. *Heliyon*. 2018. 4: e00833.
24. Baptista A, Pinho O, Pinto E, Casal S, Mota C, et al. Characterization of protein and fat composition of seeds from common beans (*phaseolus vulgaris* L.) cowpea (*Vigna unguiculata* L. Walp) and Bambara groundnuts (*Vigna subterranean* L. Verde) from Mozambique. *Journal of Food Measurement and Characterization*. 2016. 12: 1-9.
25. Uro-Chukwu HC, Ezekwe AS, Okorie U, Eleazu CO, Uro-Chukwu FCU, et al. Hypoglycemic and Antioxidant Effects of High Concentration of Bambara Groundnut (*Vigna Subterranean*) Flour Fed on Streptozotocin-induced Diabetic Rats. *International Journal of Basic and Clinical Toxicology*. 2024. 3: 10-20.
26. Yao DN, Kouassi KN, Erba D, Scazzina F, Pellegrini N, et al. Nutritive Evaluation of the Bambara Groundnut Ci12 Landrace [*Vigna subterranea* (L.) Verdc. (Fabaceae)] Produced in Côte d'Ivoire. *International Journal of Molecular Sciences*. 2015. 16: 21428-21441.
27. Barbieri R, Coppo E, Marchese A, Daglia M, Sobar Sánchez E, et al. Phytochemicals for human disease: An update on plant-derived compounds antibacterial activity. *Microbiology Research*. 2017. 196: 44-68.
28. Saxena M, Saxena J, Nema R, Singh D, Gupta A. Phytochemistry of medicinal plants. *Journal of Pharmacognosy & Phytochemistry*. 2013. 1: 168-182.
29. Uro-Chukwu HC, Okoli EC, Okpala LC, Uro-Chukwu FNC. Antioxidant and Cytotoxic Activities of Protein Hydrolysates from Shrimp Shell Wastes, germinated soybean and pigeon pea blends: A Mixture Response Surface Methodology Approach. *Journal of Drug Delivery & Therapeutics*. 2024. 14: 7-14.
30. Iwai K, Kim M, Onodera A, Matsue H. α -Glucosidase Inhibitory and Antihyperglycemic Effects of Polyphenols in the Fruit of *Viburnum dilatatum* Thunb. *Journal of Agricultural and Food Chemistry*. 2006. 54: 4588-4592.
31. Ruzaidi A, Abbe M, Amin L, Nawalyah AG, Muhajir H, et al. Hypoglycemic properties of Malaysia cocoa (*Theobromacacao*) polyphenols-rich extract. *Int Food Res J*. 2008. 15: 305-312.
32. Esteves EA, Martino HSD, Oliveira FCE, Bressan J, Costa NMB. Chemical composition of a soybean cultivar lacking lipoxygenases (LOX2 and LOX3). *Food Chemistry*. 2010. 122: 238-242.
33. Lebiedzinska A, Szefer P. Vitamins B in grain and cereal-grain food, soyproducts and seeds. *Food Chemistry*. 2006. 95: 116-122.
34. Martino HSD, Martin BR, Weaver CM, Bressan J, Esteves EA, et al. Zinc and iron bioavailability of genetically modified soybeans in rats. *Journal of Food Science*. 2007. 72: 689-695.
35. Esteves EA, Bressan J, Costa NMB, Martin HSD, Donkin SS, et al. Modified soybean affects cholesterol metabolism in rats similarly to a commercial cultivar. *Journal of Medicinal Food*. 2011. 14: 1363-1369.
36. Uro-Chukwu HC, Ezekwe AS, Roberts F, Okari KA, Uro-Chukwu FNC. Comparative Analysis of the Anti-diabetic Effects of Cocoyam, Soya Bean & Bambara groundnut flour Fed differently to Streptozotocin (STZ)-induced Diabetic Rats. *Asian Journal of Research & Reports in Endocrinology*. 2024. 7: 60-68.
37. Duranti M, Lovati MR, Dani V, Barbiroli A, Scarafoni A, et al. The alpha' Subunit from Soybean 7S Globulin Lowers Plasma Lipids and Upregulates Liver Beta-VLDL Receptors in Rats Fed a Hypercholesterolemic Diet. *The Journal of Nutrition*. 2004. 134: 1334-1339.
38. Lee SH, Parka HJ, Chuna HK, Choa SY, Junga HJ, et al. Dietary phytic acid improves serum and hepatic lipid levels in aged ICR mice fed a high-cholesterol diet. *Nutrition Research*. 2007. 27: 505-510.
39. Kim SM, Rico CW, Lee SC, Kang MY. Modulatory Effect of Rice Bran and Phytic Acid on Glucose Metabolism in High Fat-Fed C57BL/6N Mice. *Journal of Clinical Biochemistry and Nutrition*. 2010. 47: 12-17.
40. Teixeira Damasceno N, Apolinário E, Dias Flauzino F, Fernandes I, Abdalla D. Soy isoflavones reduce electronegative low-density lipoprotein (LDL-) and anti-LDL- autoantibodies in experimental atherosclerosis. *European Journal of Nutrition*. 2007. 46: 125-132.
41. Rios DR, Rodrigues ET, Cardoso AP, Montes MB, Franceschini SA, et al. Lack of effects of isoflavones on the lipid profile of Brazilian postmenopausal women. *Nutrition*. 2008. 24: 1153-1158.
42. Uro-Chukwu HC, Ezekwe AS, Roberts F, Okari KA, Uro-Chukwu FCU. Biochemical Effects of High Concentration of Colocasia Esculenta Flour Fed on Streptozotocin-Induced Diabetic Rats. *International Research Journal of Gastroenterology & Hepatology*. 2024. 7: 44-60.
43. Saito K, Jin DH, Ogawa T, Muramoto K, Hatakeyama E, et al. Antioxidative Properties of Tripeptide Libraries Prepared by the Combinatorial Chemistry. *Journal of Agricultural and Food Chemistry*. 2003. 51: 3668-3674.
44. Takahashi T, Nakamura A, Kato M, Maeda H, Mandella RC, et al. Soluble soybean fiber: a 3-month dietary toxicity study in rats. *Food Chem Toxicol*. 2003. 41: 1111-1121.
45. Takenaka Y, Doyama N, Maruyama N, Utsumi S, Yoshikawa M. Introduction of DPR, an enterostatin fragment peptide, into soybean beta-conglycinin alpha' subunit by site-directed mutagenesis. *Bioscience, Biotechnology, and Biochemistry*. 2004. 68: 253-256.
46. Ascencio C, Torres N, Isoard-Acosta F, Gímez-Pérez FJ, Hernández-Pando R, et al. Soy Protein Affects Serum Insulin and Hepatic SREBP-1 mRNA and Reduces Fatty Liver in Rats. *The Journal of Nutrition*. 2004. 134: 522-529.
47. NRC (National Research Council). Guide for the care and use of laboratory Animals. Bethesda (MD): National Institute of Health. 1985. 8523.

48. Nair AB, Jacob S. A simple practice guide for dose conversion between animals and human. *J Basic Clinical Pharmacology*. 2016. 7: 27-31.
49. Pneu-Dart. Dosage Calculation. 2023.
50. Nnadi NN, Ezekwesili CN, Ezeigwe OC. Effects of Formulated Unripe Plantain and Millet Dietary Feeds in Alloxan-Induced Diabetic Albino Rats. *International Journal of Innovative Research and Advanced Studies*. 2022. 9.
51. Yamamoto-Honda R, Takahashi Y, Mori Y, Yamashita S, Yoshida Y, et al. Changes in antidiabetic drugs prescription and glycemic control Trends in Elderly patients with type 2 diabetes mellitus from 2005-2013: An analysis of the National Centre for Diabetes Database. *Internal Medicine*. 2018. 57.
52. Azeez TA. The pattern of antidiabetic drugs and glycemic control among type 2 diabetes patients in an endocrinology clinic in Lagos, Nigeria. 2023.
53. Uro-Chukwu HC. Anti-hyperglycemic Effects of Cocoyam (*Colocasia esculenta*)-Bambara Groundnut (*Vigna subterranean*)-Soya bean (*Glycine max. (L.) Merrill*) (CY-SB-BGN) Flour Blend Fed on Streptozotocin-induced Diabetic Rats, a Ph.D Dissertation submitted to the Department of Medical Biochemistry, Rivers State University, Nkporlu, Port Harcourt, Nigeria. 2024.
54. Huang DW, Shen SC. Caffeic acid and cinnamic acid ameliorate glucose metabolism via modulating glycogenesis and gluconeogenesis in insulin-resistant mouse hepatocytes. *Journal of Functional Foods*. 2012. 4: 358-366.
55. Oboh G, Ogunbadejo MD, Ogunsuyi OB, Oyeleye SI. Can gallic acid potentiate the antihyperglycemic effect of acarbose and metformin? Evidence from streptozotocin-induced diabetic rat model. *Archives of Physiology and Biochemistry*. 2020.
56. Oršolić N, Sirovina D, Odeh D, Gajski G, Balta V, et al. Efficacy of caffeic acid on diabetes and its complications in the mouse. 2021. 26: 3262.
57. Huang DW, Chang WC, Wu JS, Shih RW, Shen SC. Gallic acid ameliorates hyperglycemia and improves hepatic carbohydrate metabolism in rats fed a high-fructose diet. *Nutrition Research*. 2016. 36: 150-60.
58. Kahkeshani N, Farzaei F, Fotouhi M, Alavi SS, Bahramsoltani R, et al. Pharmacological effects of gallic acid in health and diseases: a mechanistic review. *Iran Journal Basic Medical Science*. 2019. 22: 225-237.
59. Boaduo NK, Katerere D, Eloff JN, Naidoo V. Evaluation of six plant species used traditionally in the treatment and control of diabetes mellitus in South Africa using in vitro methods. *Pharmaceutical Biology*. 2014. 52: 756-761.
60. Garba HA, Mohammed A, Ibrahim MA, Shuaibu MN. Effect of lemongrass (*Cymbopogon citratus* Stapf) tea in a type 2 diabetes rat model. *Clin. Phytosci*. 2020. 19.
61. Kim KE, Ko KH, Heo RW, Yi CO, Shin HJ. et al. *Artemisia annua* leaf extract attenuates hepatic steatosis and inflammation in high-fat diet-fed mice. *Journal of Medical Food*. 2016. 19: 290-299.
62. Xiang M, Chen Z, He L, Xiong G, Lu J. Transcription profiling of artemisinin-treated diabetic nephropathy rats using high-throughput sequencing. *Life Science*. 2019. 219: 353-363.
63. Li X, Watanabe K, Kimura I. Gut Microbiota Dysbiosis Drives and Implies Novel Therapeutic Strategies for Diabetes Mellitus and Related Metabolic Diseases. *Frontiers in Immunology*. 2017. 8: 1882.
64. Guo Y, Fu W, Xin Y, Bai J, Peng H, et al. Antidiabetic and antiobesity effects of artemether in db/db mice. *BioMed Research International*. 2018. 8: 39-52.
65. Han P, Wang Y, Zhan H, Weng W, Yu X, et al. Artemether ameliorates type 2 diabetic kidney disease by increasing mitochondrial pyruvate carrier content in db/db mice. *American Journal of Translational Research*. 2019. 11: 1389-1402.
66. Lai CS, Tsai ML, Badmaev V, Jimenez M, Ho CT, et al. Xanthigen suppresses preadipocyte differentiation and adipogenesis through down-regulation of PPAR γ and C/EBPs and modulation of SIRT-1, AMPK, and FoxO pathways. *Journal of Agricultural Food Chemistry*. 2012. 60: 1094-1101.
67. Yu L, Chen JF, Shuai X, Xu Y, Ding Y, et al. Artesunate protects pancreatic beta cells against cytokine-induced damage via SIRT1 inhibiting NF- κ B activation. *Journal of endocrinological investigation*. 2016. 39: 83-91.
68. Ben-Othman N, Vieira A, Courtney M, Record F, Gjernes E, et al. Long-term GABA administration induces alpha cell-mediated beta-like cell neogenesis. *Cell*. 2017. 168: 73-85.
69. Vieira A, Ben-Othman N, Collombat P. GABA triggers pancreatic β -like cell neogenesis. *Cell Cycle*. 2017. 16: 727-728.
70. Holla BS, Poorjary KN, Rao BS, Shivananda MK. New bis-aminomercaptotriazoles and bis-triazolothiadiazoles as possible anticancer agents. *European Journal Medical Chemistry*. 2002. 37: 511-517.
71. Yousif E, Majeed A, Al-Sammarrae K, Salih N, Salimon J, et al. Metal complexes of Schiff base: preparation, characterization and antibacterial activity. *Arabian Journal of Chemistry*. 2013. 5: S1639-S1644.
72. Upadhyay PK, Mishra P. "Synthesis antimicrobial and anticancer activities of 5-(4 substituted phenyl) 1,3,4 thiadiazole 2 amines". *Rasayan Journal of Chemistry*. 2017. 10: 254-262.
73. Ibrahim SI, Ameh DA, Atawodi SE, Umar IA. Carbonic Anhydrase: A New Therapeutic Target for Managing Diabetes. *Journal of Metabolic Syndrome*. 2016. 5: 196.
74. Ghosh AK, Brindisi M. Urea Derivatives in Modern Drug Discovery and Medicinal J. *Med. Chem*. 2020. 63: 2751-2788.
75. Garuti L, Roberti M, Bottegoni G, Ferraro M. Diaryl Urea: A Privileged Structure in Anticancer Agents. *Curr Med Chem*. 2016. 23: 1528-1548.
76. Ullah I, Hassan M, Khan KM, Sajid M, Umar M, et al. Thiourea derivatives inhibit key diabetes-associated enzymes and advanced glycation end-product formation as a treatment for diabetes mellitus. *International Union of Biochemistry and Molecular Biology Life*. 2022. 75: 161-180.
77. Furukawa A, Arita T, Fukuzaki T, Mori M, Honda T, et al. Synthesis and biological evaluation of novel (-) cercosporamide derivatives as potent selective PPAR γ modulators. *Bioorg Med Chem Lett*. 2012. 54: 522-533.

78. Cho SK, Huh JH, Yoo JS, Kim JW, Lee KJ. HOMA-estimated insulin resistance as an independent prognostic factor in patients with acute pancreatitis. *Scientific Report*. 2019. 9: 14894.
79. Chari ST, Zapiach M, Yadav D, Rizza RA. Beta-cell function and insulin resistance evaluated by HOMA in pancreatic cancer subjects with varying degrees of glucose intolerance. *Pancreatology*. 2005. 5: 229-233.
80. Snehalatha C, Mary S, Selvam S, Sathish Kumar CK, Shetty SB, et al. Changes in insulin secretion and insulin sensitivity in relation to the glycaemic outcomes in subjects with impaired glucose tolerance in the Indian Diabetes Prevention Programme-1 (IDPP-1). *Diabetes Care*. 2009. 32: 1796-1801.
81. Katsuki A, Sumida Y, Gabazza EC, Murashima S, Furuta M, et al. Homeostasis Model Assessment is a Reliable Indicator of Insulin Resistance During Follow-up of Patients with Type 2 Diabetes. *Diabetes Care*. 2001. 24: 362-365.
82. Garg MK, Dutta MK, Mahalle N. Study of beta-cell function (by HOMA model) in metabolic syndrome. *Indian J Endocrinol Metab*. 2011. 15: S44-S49.
83. Appleton DJ, Rand JS, Sunvoid GD. Basal Plasma Insulin and Homeostasis Model Assessment (HOMA) are indicators of Insulin Sensitivity in Cats. *Journal of Feline Medicine & Surgery*. 2016. 7: 183-193.
84. Sung K, Reaven GM, Kim SH. Utility of Homeostasis Model Assessment of β -Cell Function in Predicting Diabetes in 12,924 Healthy Koreans: Response to Boyko. *Diabetic Care*. 2010. 33: 200-202.
85. Haffner SM, Kennedy E, Gonzalez C, Stern MP, Miettinen H. A prospective analysis of the HOMA model: the Mexico City Diabetes Study. *Diabetes Care*. 1996. 19: 1138-1141.
86. Song Y, Manson JE, Tinker L, Howard BV, Kuller LH, et al. Insulin sensitivity and insulin secretion determined by homeostasis model assessment and risk of diabetes in a multi-ethnic cohort of women: the Women's Health Initiative Observational Study. *Diabetes Care*. 2007. 30: 1747-1752.
87. Wallace TM, Levy JC, Matthews DR. Use and abuse of HOMA modelling. *Diabetes Care*. 2004. 27: 1487-1495.
88. Uro-Chukwu HC, Ezekwe AS, Eleazu C, Okorie UC, Uro-Chukwu FC. Body Weight Changes, Histological Features and Anti-hyperglycemic Effects of Cocoyam-Soya Bean-Bambara Groundnut Flour Blend-Fed Streptozotocin (STZ)-induced Diabetic Rats. *London Journal of Medical & Health Research*. 2024. 24: 9-25.
89. Ram H, Kumar P, Purohit A, Kashyap P, Kumar S, et al. Improvements in HOMA indices and pancreatic endocrinal tissues in type2- diabetic rats by DPP-4 inhibition and antioxidant potential of an ethanol fruit extract of withania coagulans. *Nutrition & Metabolism*. 2021. 18: 43.
90. Kitada M, Koya D. SIRT1 in type 2 diabetes: mechanisms and therapeutic potential. *Diabetes Metabolism Journal*. 2013. 37: 315-325.